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# Computational evaluation of 2-arylbenzofurans for their potential use against SARS-CoV-2: A DFT, molecular docking, molecular dynamics simulation study

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Natural compounds obtained from various sources have been used in the treatment of many diseases for many years and are very important compounds for drug development studies. They can also be an option to treat COVID-19, which is affecting the whole world and not curable with medication, yet. In this study, two 2-arylbenzofuran derivatives from *Sesbania cannabina* which are newly entered the literature were investigated computationally with the assistance of computational techniques including DFT calculations, molecular docking calculation and molecular dynamics simulations. The study consists of four parts, in the first part of the study DFT calculations were performed on the 2-arylbenzofurans, and geometry optimizations, vibrational analyses, molecular electrostatic potential (MEP) map calculations, frontier molecular orbital (FMO) calculations and Mulliken charge analyses were carried out. In the second part, molecular docking calculations were performed to investigate the interactions between the molecules and two potential target, SARS-CoV-2 main protease (SARS-CoV-2 M<sup>pro</sup>) and SARS-CoV-2 spike receptor binding domain – human angiotensin converting enzyme 2 complex (SARS-CoV-2 SRBD – hACE2). In the third part, MD simulations were performed on the top-scoring ligand – receptor complexes to investigate the stability of the complex and the interactions between ligands and receptors in more detail. Finally, drug-likeness analyses and ADME (adsorption, desorption, metabolism, excretion) predictions were performed on the investigated compounds. Results showed that investigated natural compounds effectively interacted with the target receptors and gave comparable results to the reference drug molecules.

Keywords: ACE2, COVID-19, Drug-likeness, In silico investigation, SARS-CoV-2 main protease

SARS-CoV-2 continues to spread rapidly all over the world since its first appearance in December 2019 and grew into a global pandemic nowadays. Although there are many studies carried out all over the world against SARS-CoV-2, an effective and specific drug treatment has not yet been developed. Studies against COVID-19 can be divided into three groups: (1) vaccine development, (2) drug repurposing and (3) new drug development. Since some drugs on the market are known to be beneficial in the treatment of COVID-19, they are used in the treatment of this disease. However, these drugs are not the drugs developed specifically for COVID-19 and their effectiveness is not at the desired level. For now, although vaccine development studies seem to be the most effective way to develop a treatment in a short time, development of new specific drugs is needed and will be needed in the future, too.

SARS-CoV-2 M<sup>pro</sup> has an important role in viral replication and transcription, and this makes it an appealing target for the treatment of COVID-19. On the

other hand, it is known that SARS-CoV-2 uses its spike receptor binding domain to bind to human angiotensin converting enzyme-2 (hACE2) and enter the human cell. Preventing this interaction between SARS-CoV-2 SRBD and hACE2 could provide an important method for treating the disease, as it could prevent the entrance of the virus into the human cell. In the literature, there are various studies targeted the inhibition of SARS-CoV-2 M<sup>pro1-6</sup> or preventing SARS-CoV-2 SRBD to interact with hACE2<sup>7-9</sup>.

In this study, two natural compounds which have recently been added to the literature by Fu *et al*<sup>10</sup>, have been investigated computationally. Fu *et al.* have evaluated the anticancer activities of these natural compounds in their study but to the best of our knowledge these compounds have not yet been studied experimentally or computationally for their antiviral effects against SARS-CoV-2. The structures of these 2-arylbenzofuran derivatives are given in (Fig. 1)<sup>10</sup>.

# **Materials and Methods**

#### **DFT Calculations**

In this part of the study, geometry optimizations, vibrational analyses, MEP map calculations, FMO

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Fig. 1 — Molecular structures of 2-arylbenzofurans

calculations and Mulliken charge analyses were carried out. All calculations in this part were performed in both gas and solution phasewith the use of Becke threeparameter hybrid functional combined with Lee-Yang-Parr correlation functional (B3LYP) and Austin-Frisch-Petersson functional with dispersion method (APF-D). In the calculations performed in solution phase, IEFPCM (Polarizable Continuum Model using the integral equation formalism variant) solvation model was used, and water was selected as solvent. All the calculations were performed with the use 6-311+G(2d,p) basis set. Gaussian 09 Rev.D01<sup>11</sup>, GaussView 5<sup>12</sup>, VeraChemV conf<sup>13</sup> software packages were used in the calculations.

# **Molecular Docking Calculations**

In molecular docking calculations, the structures of SARS-CoV-2 M<sup>pro</sup>and SARS-CoV-2 SRBD - hACE2 were obtained from RCSB Protein Data Bank (PDB ID: 5r80 and  $6lzg)^{14-16}$ . The 3D structures of 2-arylbenzofurans were obtained from DFT calculations. In molecular docking calculations, AutoDock Tools<sup>17</sup> and AutoDock Vina<sup>18</sup> program packages were used and Discovery Studio Visualizer<sup>19</sup> was used in the representation of the docking results. Water and ligand molecules were removed, hydrogens and Gasteiger charges were added to prepare the receptors for docking. Lamarckian genetic algorithm was used in the docking procedure. Gridbox size was set to  $20 \times 25 \times 20$  Å<sup>3</sup> and  $20 \times 20 \times 20$  Å<sup>3</sup> for SARS-CoV-2 M<sup>pro</sup> and SARS-CoV-2 SRBD - hACE2, respectively, and grid spacing was set to 1 Å in all cases. After each docking calculation, nine docking poses having highest binding scores were obtained.

## **Molecular Dynamics Simulations**

MD simulation studies on top-scoring ligand – receptor complexes followed the molecular docking calculations to investigate the interactions in more detail and to investigate the stability of the complexes. MD simulations were performed for 30 ns. In MD simulations, GROMACS 2020<sup>20</sup> program package was used. Ligand topologies were obtained from Acpype Server<sup>21</sup> and topologies of the receptors were obtained with the use of AMBER<sup>22</sup> force field and



Fig. 2 — Geometry optimized structures of the investigated compounds

TIP3P water model. First, in each case, system was neutralized, and energy minimization procedure was run by employing steepest descent minimization algorithm. NVT and NPT ensemble equilibrations were performed for 200 ps. MD simulations were performed in a dodecahedron simulation box at 1 bar and 300 K reference pressure and temperature.

# **Drug-likeness Analyses and ADME Predictions**

Drug-likeness analyses and ADME predictions were performed *via* Swiss ADME<sup>23</sup>web server and some important parameters such as physicochemical properties, lipophilicity and water solubility were predicted. In this part, Lipinski<sup>24</sup>, Ghose<sup>25</sup>, Veber<sup>26</sup>, Egan<sup>27</sup> and Muegge<sup>28</sup> filters were used to perform drug-likeness analysis on the investigated compounds.

#### **Results and Discussion**

#### **DFT Calculations**

In this part, a conformer search was carried out prior to geometry optimizations in order to assure that global minimum was obtained for each structure. Additionally, frequency calculations followed the geometry optimizations to confirm that each optimized geometry corresponds to a global minimum. Geometry optimized structures of the investigated compounds which were obtained in gas phase with the use of B3LYP method and 6-311+G(2d,p) basis set are given in (Fig. 2). In Figure 3, calculated infrared spectra of the investigated compounds obtained at the same level of theory are given.

MEP maps give information about the electron rich and electron deficient parts of a given molecule. These electron rich and electron deficient centers play important role in the interactions between investigated



Fig. 3 — Calculated infrared spectra of the investigated compounds (A) compound 1; and (B) compound 2

compounds and receptors. MEP maps of the investigated compounds are given in (Fig. 4). Results showed that negative charge was predominantly on the oxygen atoms, while hydroxyl group hydrogens carried positive charge. The results also showed that the method used in the calculations and the phase in which the calculations were performed (gas or solution phase) affect the charge distribution in MEP maps. It was observed that the charge ranges obtained from the calculations performed in solution phase are wider than the ranges obtained from the calculations performed in gas phase. It was also seen that the charge ranges obtained with the use of APF-Dare wider than the ranges obtained with the use of B3LYP.

HOMO and LUMOs of a given molecule are used to estimate how this molecule interacts with other compounds. HOMO and LUMOs of the investigated compounds and the energies obtained with the use of different methods are given in (Fig. 5). Results showed that in gas phase, HOMO-LUMO gaps of compound 2 were found to be larger than those of compound 1. In contrast to gas phase, results obtained from the calculations performed in water showed that HOMO-LUMO gaps of compound 1 were found to be larger than those of compound 2. FMO analyses indicated that delocalization is occurred over all atoms except methyl groups for both molecules. Additionally, relatively large HOMO-LUMO gaps indicate that both molecules have quite stable structures.

Calculated Mulliken charges of the investigated compounds with the use of B3LYP and APF-D in gas phase and solution phase are given in (Table 1). In both cases 6-311+G(2d,p) basis set was used.



Fig. 4 — MEP maps of the investigated compounds



Fig. 5 - FMOs of the investigated compounds

Mulliken atomic charges of a given molecule significantly influence some important parameters such as dipole moment, polarizability, and electronic structure. For both compounds, all hydrogen atoms have a positive charge while all oxygen atoms have a negative charge. On the other hand, the Mulliken charges of carbon atoms highly influenced by their environment and have both positive and negative charges.

# **Molecular Docking Calculations**

Molecular docking calculations were performed to obtain top-scoring docking poses for compounds 1 and 2. Additionally, to make a comparison, calculations were repeated for remdesivir, lopinavir, favipiravir and hydroxychloroquine which are the drugs currently under investigation for their potential use against COVID-19. Especially remdesivir is widely used because of being an FDA-approved drug for the treatment of COVID-19. 3D structures of remdesivir, favipiravir and hydroxychloroquine were obtained from PubChem Database<sup>29</sup> and the structure of lopinavir was obtained from Zinc Database<sup>30</sup>. Results for SARS-CoV-2-SRBD – hACE2 showed that, investigated 2-arylbenzofurans have higher binding score than all other compounds. On the other hand, for SARS-CoV-2 M<sup>pro</sup>, the binding scores of 2-arylbenzofurans are higher than those of favipiravir and hydroxychloroquine and comparable to the binding scores of lopinavir and remdesivir (Table 2).

# **Molecular Dynamics Simulations**

After performing molecular docking calculations, 30 ns molecular dynamics simulations were performed for each top-scoring ligand-receptor complex to investigate the stability and the ligandreceptor interactions in more detail.

In Figure 6, the superimposed structures of SARS-CoV-2  $M^{pro}$  – 1 complex extracted every 5 ns of the simulation are given. 2D representations of these interactions are given in (Fig. 7). Results showed that ligand remained in the receptor binding pocket throughout the entire simulation.

		Tr	able 1 — N	Iulliken atomic c	harges of cor	nnounds 1 a	and 2		
		Mulliken			indiges of col	npoundo i e		h Charges	
Comp. 1	B3LYP	B3LYP/H <sub>2</sub> O	APF-D	APF-D/H <sub>2</sub> O	Comp. 2	B3LYP	B3LYP/H <sub>2</sub> O	APF-D	APF-D/H <sub>2</sub> O
1C	-0.286	-0.345	-0.427	-0.474	1C	-0.061	-0.055	0.054	0.078
2C	0.382	0.391	0.347	0.345	2C	0.319	0.306	0.255	0.223
2C 3C	-0.481	-0.425	-0.482	-0.437	2C 3C	-0.242	-0.257	-0.424	-0.431
3C 4C	0.249	0.254	0.158	0.148	4C	0.242	0.169	-0.010	-0.030
5C	0.249	0.696	0.138	0.928	4C 5C	0.721	0.704	0.920	0.925
5C 6C	-0.254	-0.276	-0.346	-0.370	5C 6C	-0.754	-0.736	-0.897	-0.930
0C 7O	-0.454	-0.486	-0.388	-0.418	70	-0.570	-0.575	-0.502	-0.506
70 8C	-0.158	-0.168	-0.252	-0.266	70 8C	-0.148	-0.165	-0.245	-0.264
90	-0.400	-0.416	-0.353	-0.371	90	-0.410	-0.438	-0.369	-0.400
10C	0.400	-0.003	0.083	-0.039	10C	0.410	0.269	0.30)	0.311
10C 11C	0.402	0.313	0.636	0.549	10C 11C	0.270	0.393	0.764	0.708
11C 12C	0.063	0.208	-0.158	0.017	11C 12C	0.201	0.367	-0.048	0.159
130	-0.419	-0.508	-0.405	-0.487	130	-0.414	-0.501	-0.402	-0.481
140	-0.321	-0.364	-0.292	-0.336	140	-0.326	-0.368	-0.297	-0.342
140 15C	0.288	0.224	0.292	0.245	140 15C	0.328	0.308	0.297	0.342
15C 16C	0.288	0.224	0.231	0.245	15C 16C	0.328	0.432	0.442	0.310
10C 17C	-0.499	-0.440	-0.448	-0.426	10C 17C	-0.516	-0.450	-0.476	-0.442
17C 18C	0.040	0.056	0.062	0.005	17C 18C	0.057	0.430	0.096	0.031
18C 19C	-0.286	-0.276	-0.367	-0.333	18C 19C	-0.327	-0.309	-0.438	-0.390
20C	0.322	0.276	0.262	0.249	20C	0.327	0.351	0.438	0.390
200	-0.460	-0.505	-0.454	-0.500	200	-0.455	-0.504	-0.448	-0.498
210	-0.429	-0.467	-0.413	-0.448	210	-0.429	-0.466	-0.413	-0.447
220	-0.442	-0.466	-0.351	-0.379	220	-0.440	-0.464	-0.345	-0.376
230 24C	-0.157	-0.168	-0.267	-0.260	230 24C	-0.157	-0.167	-0.272	-0.263
24C 25H	0.094	0.112	0.114	0.133	250	-0.478	-0.527	-0.477	-0.522
26H	0.100	0.112	0.114	0.133	250 26H	0.088	0.120	0.109	0.141
2011 27H	0.100	0.128	0.119	0.147	2011 27H	0.088	0.120	0.109	0.141
2711 28H	0.100	0.120	0.124	0.142	2711 28H	0.139	0.117	0.120	0.140
29H	0.139	0.151	0.160	0.172	29H	0.139	0.155	0.161	0.175
30H	0.130	0.152	0.162	0.177	30H	0.142	0.155	0.164	0.173
31H	0.138	0.132	0.102	0.292	31H	0.245	0.134	0.249	0.293
32H	0.243	0.289	0.230	0.232	32H	0.243	0.290	0.249	0.138
33H	0.071	0.117	0.100	0.138	33H	0.071	0.117	0.101	0.135
34H	0.081	0.329	0.100	0.333	34H	0.081	0.330	0.101	0.333
34H 35H	0.288	0.329	0.292	0.333	34H 35H	0.290	0.313	0.294	0.333
35H 36H	0.273	0.156	0.280	0.176	35H 36H	0.273	0.156	0.279	0.319
30H 37H	0.144	0.150	0.103	0.176	30H 37H	0.144	0.150	0.184	0.176
37H 38H	0.133	0.159	0.188	0.180	37H 38H	0.133	0.159	0.188	0.180
5011	0.150	0.133	0.14/	0.1/3	39H	0.131	0.133	0.147	0.173
					5911	0.310	0.343	0.327	0.555

Table 2 — Binding sc	ores of the investiga	ted compounds.
Compound	6LZG <sup>1</sup>	5R80 <sup>2</sup>
1	-7.5	-7.6
2	-7.4	-7.0
Remdesivir	-6.9	-7.9
Lopinavir	-6.8	-8.2
Favipiravir	-6.8	-5.1
Hydroxychloroquine	-5.9	-6.3
<sup>1</sup> SARS-CoV-2 SRBD – hA <sup>2</sup> SARS-CoV-2 M <sup>pro</sup>	ACE2	

Results for SARS-CoV-2 M<sup>pro</sup> – 1 complex showed that, in most of the simulation time, MET49, PRO168, MET165, GLN189, HIS41, THR190 and ARG188 took part in the interactions between ligand and receptor.

Superimposed structures of the SARS-CoV-2  $M^{pro}$  – 2 complex extracted every 5 ns of the simulation are given in (Fig. 8) and 2D representations of these interactions are given in (Fig. 9). Results showed that ligand remained in the receptor binding pocket throughout the entire simulation.



Fig. 6 — Superimposed structures of SARS-CoV-2  $M^{pro}$  – 1 complex extracted every 5 ns of the simulation

Results showed that, when compared to SARS-CoV-2  $M^{pro} - 1$  complex, for SARS-CoV-2  $M^{pro} - 2$  complex there were considerable differences in the interactions between ligand and receptor. In this complex, in most of the simulation time, CYS145, THR26, SER46 and THR25 took part in the interactions between ligand and receptor. When considered together with the binding score, it can be concluded that the interactions in this complex are lower than those in the SARS-CoV-2  $M^{pro} - 1$  complex.

Superimposed structures of SARS-CoV-2 SRBDhACE2 - 1 complex extracted every 5 ns of the



Fig. 7 - 2D representations of the ligand - receptor interactions in SARS-CoV-2 M<sup>pro</sup>-1 complex

simulation are given in (Fig. 10) and 2D representations of these interactions are given in (Fig. 11). Results showed that ligand remained in the receptor binding pocket in the interface between SARS-CoV-2 SRBD and hACE2 throughout the entire simulation.



Fig. 8 — Superimposed structures of SARS-CoV-2  $M^{pro}$  – 2 complex extracted every 5 ns of the simulation

Results for SARS-CoV-2 SRBD-hACE2 – 1 complex showed that, throughout almost the entire simulation period, PRO389, ASP30 and HIS34 took part in the interactions between ligand and receptor. Additionally, LEU29, GLN96, LYS417, LYS26, ASN33, GLN388,



Fig. 10 — Superimposed structures of SARS-CoV-2 SRBDhACE2 – 1 complex extracted every 5 ns of the simulation



Fig. 9 — 2D representations of the ligand – receptor interactions in SARS-CoV-2 M<sup>pro</sup>- 2 complex

ARG393, ALA386, ALA387, ARG403 and TYR453 took part in the stabilization of the complex.

Finally, superimposed structures of SARS-CoV-2 SRBD-hACE2 – 2 complex extracted every 5 ns of the simulation are given in (Fig. 12 and 2D) representations of these interactions are given in (Fig. 13). Results showed that ligand remained in the receptor binding pocket in the interface between SARS-CoV-2 SRBD and hACE2 throughout the entire simulation.

Results showed that, when compared to SARS-CoV-2 SRBD-hACE2 - 1 complex, for SARS-CoV-2 SRBD-hACE2 - 2 complex there were some differences in the interactions between ligand and receptor. In this complex, in most of the simulation time, HIS34,

GLU35, SER494, TYR505, ARG403, GLU406 and ASN33 took part in the interactions between ligand and receptor. Additionally, GLU37, LYS353, ARG393, ALA387, PRO389, TYR495, ASP38 and GLN388 participated in the interactions.



Fig. 12 — Superimposed structures of SARS-CoV-2 SRBDhACE2 – 2 complex extracted every 5 ns of the simulation



Fig. 11 — 2D representations of the ligand – receptor interactions in SARS-CoV-2 SRBD-hACE2 – 1 complex

The interactions in the structures of all complexes belong to  $30^{\text{th}}$  ns of the simulation are shown in (Fig. 14).

Root mean square deviation (RMSD) and radius of gyration (RG) are often used to determine the stability of proteins. RMSDs of proteins compared to their initial positions in all four ligand-receptor complexes are given in (Fig. 15). Average RMSD values of proteins and standard deviations were found to be 0.2535 (0.1306),

0.2491 (0.1321), 0.2548 (0.1297) and 0.2954 (0.1346) nm for SARS-CoV-2  $M^{pro}$  - 1, SARS-CoV-2  $M^{pro}$  - 2, SARS-CoV-2 SRBD-hACE2 - 1 and SARS-CoV-2 SRBD-hACE2 - 2 complexes, respectively.

In the study, RMSDs of ligands compared to the position of the receptors were monitored for each complex to investigate how well the binding poses were preserved during the MD simulation. Obtained results are given in (Fig. 16).



Fig. 13 — 2D representations of the ligand – receptor interactions in SARS-CoV-2 SRBD-hACE2 – 2 complex

Results showed that there was no considerable change in the relative position of the ligand in SARS-CoV-2  $M^{pro}$  1 and SARS-CoV-2 SRBD-hACE2 – 2 complexes (Fig. 16A-D). For SARS-CoV-2 M^{pro} – 2 and SARS-CoV-2 SRBD-hACE2 – 1 complexes, it was found that the position of the ligand reached its equilibrium position about 15<sup>th</sup> ns and after this time there was no considerable change in the relative position of the ligand.

Radius of gyration is another parameter to estimate the stability of a given protein. RG of the proteins in all four complexes were monitored during the MD simulation and are given in (Fig. 17).

Average RG values of proteins and standard deviations were found to be 2.2316 (0.0138), 2.2331 (0.0141), 3.1341 (0.0204)and 3.1296 (0.0205) nm for SARS-CoV-2  $M^{pro} - 1$ , SARS-CoV-2  $M^{pro} - 2$ , SARS-CoV-2 SRBD-hACE2 – 1 and SARS-CoV-2 SRBD-



Fig. 14 — The interactions in the structures of the complexes belong to  $30^{\text{th}}$  ns of the MD simulation (A) SARS-CoV-2 M<sup>pro</sup>- 1; (B) SARS-CoV-2 M<sup>pro</sup>- 2; (C) SARS-CoV-2 SRBD-hACE2 - 1; and (D) SARS-CoV-2 SRBD-hACE2 - 2



Fig. 15 — RMSDs of proteins compared to their initial positions (A) SARS-CoV-2 M<sup>pro</sup>-1; (B) SARS-CoV-2 M<sup>pro</sup>-2; (C) SARS-CoV-2 SRBD-hACE2 - 1; and (D) SARS-CoV-2 SRBD-hACE2 - 2

hACE2 - 2 complexes, respectively. Results showed that there was no considerable change in the radius of gyration of the proteins in all of the complexes.

# **Drug-likeness and ADME Predictions**

In the study, in addition to DFT calculations, molecular docking calculations and molecular dynamics simulations, drug-likeness analyses and ADME predictions were performed via Swiss ADME web server<sup>23</sup>. Drug-likeness and ADME prediction results are given in (Tables 3 & 4).

Results showed that both molecules have no more than 10 hydrogen bond acceptors and 5 hydrogen bond donors which means both molecules obey the Lipinski rule. Molecular weights do not exceed the



Fig. 16 — RMSDs of ligands compared to the positions of the receptors (A) SARS-CoV-2  $M^{pro}$  - 1; (B) SARS-CoV-2  $M^{pro}$  - 2; (C) SARS-CoV-2 SRBD-hACE2 - 1; and (D) SARS-CoV-2 SRBD-hACE2 - 2



Fig. 17 — Change in the RG of protein during the MD simulation (A) SARS-CoV-2  $M^{pro} - 1$ ; (B) SARS-CoV-2  $M^{pro} - 2$ ; (C) SARS-CoV-2 SRBD-hACE2 – 1; and (D) SARS-CoV-2 SRBD-hACE2 – 2

Table 3 — Results of investigated	ADME analyses	of the
Compound	1	2
Physicochemical Properties		
Number of H-Bond	7	8
Acceptors		
Number of H-Bond Donors	3	4
Molecular Weight (g/mol)	330.3	346.3
Number of heavy atoms	24	25
Number of aromatic heavy atoms	15	15
Fraction C sp <sup>3</sup>	0.12	0.12
Number of rotatable bonds	4	4
Molar refractivity	85.64	87.66
TPSA ( $Å^2$ )	109.36	129.59
Lipophilicity		
iLOGP	2.26	1.95
XLOGP3	2.75	2.39
WLOGP	3.23	2.93
MLOGP	0.78	0.26
SILICOS-IT	2.45	1.97
Consensus logP <sub>o/w</sub>	2.29	1.90
Water Solubility		
ESOL	S	S
ALI	MS	MS
SILICOS-IT	MS	S
Pharmacokinetics		
GI Absorption	High	High
BBB Permeation	No	No
Skin permeation, logK <sub>p</sub> (cm/s)	-6.36	-6.72

TPSA: Topological Polar Surface Area, GI: Gastrointestinal, BBB: Blood Brain Barrier, MS: Moderately soluble, S: Soluble, VS: Very soluble

limits suggested by Lipinski and Ghose rules. Number of rotatable bonds was found to be 4 for both molecules and met the requirements of Veber and Muegge rules. Fraction C sp<sup>3</sup> value was found to be 0.12 for both molecules. Increasing fraction C sp<sup>3</sup> values correspond to higher degrees of saturation and it is proposed that increasing degrees of saturation increases the clinical success rate of a molecule<sup>31,32</sup>.

Molar refractivities of the molecules 1 and 2 were found to be 85.64 and 87.66, respectively. Since the Ghose rule states that the molar refractivity should be between 40-130, both molecules fulfill this rule. In prediction of the drug-likeness of a given molecule, topological polar surface area (TPSA) is also used<sup>33</sup>. TPSA values of the compounds were found to be 109.36 and 129.59 Å<sup>2</sup>, respectively, and met the requirements of Veber rule. As is known, Veber rule

	mistry analyses	2
Compound	I	2
Drug-likeness		
Lipinksi	Yes	Yes
Ghose	Yes	Yes
Veber	Yes	Yes
Egan	Yes	Yes
Muegge	Yes	Yes
ABS	0.56	0.56
Medicinal Chemistry		
PAINS	0 alert	0 alert
Brenk	0 alert	0 alert
Leadlikeness	Yes	Yes
SAS	3.32	3.44
ABS: Abbott Bioavailabilty	Score PAINS Pan A	6691/

stated that the TPSA of a given molecule should not be greater than 140 Å<sup>2</sup>. The average partition coefficient<sup>34-40</sup> of the molecules 1 and 2 were found to be 2.29 and 1.90, respectively. According to ESOL<sup>41</sup> and ALI<sup>42</sup>, water solubilities of molecules 1 and 2 were predicted to be soluble and moderately soluble, respectively. On the other hand, SILICOS-IT<sup>40</sup> predicted that the water solubility of molecules 1 and 2 are moderately soluble and soluble, respectively. Gastrointestinal absorptions of the molecules were predicted to be high, but it was predicted that molecules cannot penetrate blood-brain barrier.

In the study it was found that both molecules obey Lipinski, Ghose, Veber, Egan and Muegge rules, and bioavailability scores<sup>43</sup> were estimated to be 0.56 for both molecules. Additionally, no alert was observed for PAINS<sup>44</sup>, Brenk<sup>45</sup> and leadlikeness<sup>46</sup> analyses, and synthetic accessibility scores of molecules 1 and 2 were found to be 3.32 and 3.44, respectively.

# Conclusion

In the study, DFT calculations, molecular docking calculations, MD simulation studies, drug-likeness analyses and ADME predictions have been performed on two newly introduced 2-arylbenzofuran derivatives obtained from *Sesbania cannabina* by Fu *et al.*<sup>9</sup>, to investigate them computationally for their potential use against SARS-CoV-2. In DFT calculations, four different methods (B3LYP/6-311+G(2d,p) in gas phase, B3LYP/6-311+G(2d,p) in solution phase, APF-D/6-311+G(2d,p) in gas phase and APF-D/6-311+G(2d,p) in solution phase) were used for each compound and the results were compared. In molecular docking and MD simulation studies, two potential targets, SARS-CoV-2 M<sup>pro</sup> and SARS-CoV-2

SRBD – hACE2 complexes, suggested for the treatment of COVID-19 were selected. Results showed that the compounds were effectively bound to the active sites of the investigated receptors and remained in the binding pocket throughout the entire MD simulation. Additionally, in drug-likeness and ADME analyses, no violation was observed for the investigated 2-arylbenzofuran derivatives. In the study, it was concluded that the investigated compounds can be promising structures for further studies on COVID-19 treatment.

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# **Conflict of interest**

All authors declare no conflict of interest.

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