



BMI and adipocytokine changes in COPD exacerbation and stable COPD

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Received 11 June 2019; revised 28 April 2021

COPD is described by progressive airflow restriction and recurrent acute exacerbations, which is caused by inflammatory response occurring in lungs as a result of chronic inhalation of harmful particles and gases. The study was designed to determine the link of interleukin-6 (IL-6), adiponectin and visfatin with BMI and oxidant/antioxidant balance in Chronic Obstructive Pulmonary Disease (COPD), a systemic disease. The study comprised control (n=20), patients with COPD (n=37) from the Chest Diseases Clinic of Firat University Hospital. The serum MDA, IL-6, Visfatin and Adiponectin levels were evaluated by ELISA. Also, Pulmonary Function Tests were done. There was no significant difference between control and patients with COPD in terms of sex and age averages. On the contrary, BMI levels were statistically significantly lower in COPD group compared to the controls. MDA and adiponectin levels were higher, IL-6 and visfatin levels were lower in COPD groups contrary to controls. It was thought that the level changes of these parameters (MDA, IL-6, visfatin and adiponectin) may be an important factor in the development of COPD and in monitoring the treatment of COPD-related diseases.

Keywords: Adiponectin, Chronic obstructive pulmonary disease (COPD), Interleukin-6, Malondialdehyde (MDA), Visfatin

Chronic obstructive pulmonary disease (COPD) is a disease characterized by progressive airflow limitation and recurrent acute exacerbations, which is caused by inflammatory response occurring in lungs as a result of chronic inhalation of harmful particles and gases¹. COPD affects small and large airways in lung, lung parenchyma and pulmonary vascular bed, and also other systems².

Causes of body composition change (fat-free mass, fat mass and bone mineral content, *etc.*) accepted as an important cause of comorbidity in COPD³, the increase in energy consumption that is associated with respiration developing in parallel to the increase in respiratory act, negative energy balance caused by the impairment in metabolic and functional capacities are used to explain this change⁴. In the pathogenesis of COPD, in addition to genetic factors, environmental factors, inflammation and especially oxidative stress play an important role⁵. Activated inflammatory cells, neutrophils and alveolar macrophages lead to endogenous oxidant production in patients. Increased oxidative burden damages all the cellular macromolecules (lipid, protein and DNA)⁶.

Adiponectin, the richest adipokine in human plasma, has been reported to have both pro-inflammatory and anti-inflammatory effects^{7,8}. Therefore, while serum level of adiponectin is high in some human studies^{9,10}, it is low in others^{11,12}. Visfatin is a proinflammatory cytokine which is secreted from visceral fat tissue rather than subcutaneous fat tissue, exhibits an insulin-like effect by binding to insulin receptors and has immunomodulatory effects in inflammation, natural immunity regulation¹³. It was reported that evaluated the relationship of nesfatin and visfatin with inflammation activity, severity of symptoms, and lung function in men with emphysema and COPD¹⁴. IL-6, one of the inflammatory cytokines, increases in different lung diseases and is a by-product of ongoing inflammation in the lung¹⁵. In a study conducted by Liang *et al.*, leptin and IL-6 levels in the serum and sputum were higher in patients with acute exacerbation of COPD compared with stable COPD and control patients. Therefore, they were suggested that leptin and IL-6 levels were associated with the severity of COPD¹⁶.

It was aimed to detect the association of interleukin-6 (IL-6), adiponectin and visfatin with BMI and oxidant/antioxidant balance in COPD, a systemic inflammatory disease.

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Materials and Methods

Patient selection

37 patients who were followed with a diagnosis of COPD acute attack in the Chest Diseases Clinic of Firat University Hospital were included in the study. The diagnosis of COPD acute attack was defined as a situation which is characterized by an increase in respiratory disorder and a decrease in the daily performance, change in sputum amount and color, exacerbation in coughing, high fever and/or deterioration in mental status in a case progressing stable, and which may require change in an ordinary drug treatment with acute onset. Age, gender, duration of disease and smoking histories of the cases were questioned, and their body mass indexes were calculated by measuring weight and height.

The presence of systemic, allergic, neoplastic or immunologic diseases, the determination of reversibility in pulmonary functions and the presence of coexisting lung disease to COPD were considered as the exclusion criteria from the study. Control group was formed by randomly selecting 20 cases who were in the same age group and gender with COPD patients, who has normal spirometric lung functions and who had no history of lung and systemic infection and regular use of drug in the last month. They were also nonsmokers.

Blood was taken from COPD patients for 2 times which were in the acute period (1st day and on the 15th day after treatment, and 5 mL of blood was taken from the healthy control group for 1 times, and serum levels were studied in Medical Biochemistry Laboratory. From COPD patients and the healthy control group, venous blood samples were taken into gelled chemistry tubes between at 08:30-09:30 after 10 h of fasting, and their serums were separated by centrifuging for 5 min. at 4000 rpm. Serum samples were separated into portions and stored at -80°C until the day of analysis. Samples were studied by warming to room temperature (23–25.5°C) after taken out of the deep freezer.

Detection of Malondialdehyde (MDA)

As an indicator of lipid peroxidation, MDA levels were measured using appropriate MDA kit (Immuchrom GmbH Company, Hessen, German) by High-performance liquid chromatography (HPLC). MDA, which is converted into fluorescence products with a derivatising agent, added reaction solution. The absorbance of MDA was measured at 515 nm (excitation), at 553 nm (emission) by spectrophotometric detector. The results were given as nmol/mL.

Determination of serum levels of IL-6, Visfatin and Adiponectin

Serum concentration of IL-6 (Boster Biological Technology, Ltd., Pleasanton, CA, EK0410) Visfatin (Adipo Bioscience, Santa Clara, CA, USA, SK00121-01) and adiponectin (Boster Biological Technology, Ltd., Pleasanton, CA, EK0595) levels were measured by a commercially available sandwich enzyme-linked immunosorbent assay according to the manufacturer's protocol. Absorbance of standards and samples was read at 450 nm. Automatic ELx50 Biotek plate washer and ELx800 plate reader was used in ELISA studies. Manufacturer average intra-assay and inter-assay coefficients of variation were <10% for IL-6, visfatin and adiponectin.

Measurement of Pulmonary Function Tests

Pulmonary function tests were performed at least three measurements using nose plugs, in a sitting position, at room temperature in Firat University Chest Diseases Clinic and the best results were evaluated. FEV1, FVC and FEV1/FVC ratio were measured and recorded. The data was interpreted according to the European Respiratory Society estimates.

Statistical Analyses

All results were presented as means \pm SD and analyzed using one-way analysis of variance (ANOVA). The LSD test was used for comparisons between groups. $P \leq 0.05$ was considered statistically significant. Statistical analysis was performed using the SPSS 22.0 statistical software package (SPSS Inc., Chicago, IL, USA) for this purpose. Bivariate correlation analysis (calculation of the Pearson coefficient) was used to assess the correlation of serum visfatin and adiponectin levels to each parameter.

Results

There was no any statistical difference between the patients and the control group in terms of sex and age averages. BMI values were statistically significantly lower in COPD group compared to the controls ($P < 0.001$). Demographic features of the study group are shown in (Table 1).

Mean FEV1, FVC and FEV1/FVC values of COPD patient group (1st, 15th day and total) were quite lower

Table 1 — Demographic Characteristics of the Study Group

	Control Group (n=20)	COPD Group (n=37)
Age (year)	60.05 \pm 5.71	61.59 \pm 5.91
Sex (M/F)	20/0	37/0
BMI (kg/m ²)	23.67 \pm 1.98	21.16 \pm 2.84*

* $P < 0.001$ compared with control group

Table 2 — Pulmonary Function Test Parameters of the control and patient group with COPD (1, 15 days and total), and serum MDA, IL-6, visfatin and adiponectin levels

	Control Group	COPD Group (1 st Day)	COPD Group (15 th Day)	COPD Group (Total)
BMI	23.67±1.98	21.16±2.84*	21.63±2.89**	21.38±2.86*
FEV1 (%pred)	87.95±4.55	31.05±11.08*	39.19±13.59*	34.76±12.86*
FVC (%pred)	87.40±4.67	47.32±12.87*	55.55±17.52*	51.07±15.61*
FEV1/FVC	88.45±5.67	49.27±10.19*	50.13±11.46*	49.66±10.71*
MDA (nmol/mL)	0.64±0.22	1.61±0.24*	1.36±0.23*	1.50±0.27*
IL-6 (pg/mL)	1.23±0.42	1.11±0.08**	1.10±0.04**	1.11±0.06**
Visfatin (ng/mL)	9.71±1.36	8.37±1.26*	8.64±1.32**	8.49±1.29*
Adiponectin (ng/mL)	22.51±10.69	151.74±93.68*	162.89±104.79*	156.83±98.30*

* $P < 0.001$ compared with control group, ** $P < 0.05$ compared with control

compared to the control group ($P < 0.001$ for three parameters). Serum visfatin and IL-6 levels were statistically significant lower in COPD group compared to the controls. MDA and adiponectin levels were statistically significant higher in patient groups compared with controls (Table 2).

There was a positive correlation between serum IL-6 levels and visfatin levels ($r = 0.330$, $p = 0.046$), and a negative correlation between adiponectin ($r = -0.375$, $P = 0.022$) patient groups (Table 3).

Discussion

Decreases in bone mass in the patients with low BMI and increase in fatty tissue in obese patients were associated with the impairment in health status and increased mortality¹⁷. It was also indicated that low BMI is a risk factor for COPD development¹⁸. The reducing BMI as a result of dyspnea in COPD, inadequate food intake due to the difficulty in food digestion and excessive apoptosis in skeletal muscles based on systemic inflammation has been defined as an independent marker of mortality and morbidity secondary to the disease¹⁹. The fact that BMI values were statistically significantly lower in COPD patient group, in which all the patients are male, compared to the control group in our study supports the study results above. The relationship that we detected between BMI values and respiratory functions of the patients with COPD exacerbation gives rise to the thought that BMI assessment is important in disease pathogenesis. Moreover, understanding the weight loss mechanisms in COPD and minimizing the weight loss have been found important in terms of survival duration and fighting against the disease²⁰.

Oxidative stress and inflammation that are characteristic for COPD have synergistic effects on muscle breakdown²¹. As a result of excessive oxidant production of oxidant/antioxidant balance and/or

Table 3 — Correlation between adipokines and other parameters in patients with COPD exacerbations (n=37)

	Visfatin (ng/mL)	Adiponectin(ng/mL)
IL-6 (pg/mL)	$Rho = 0.330$ $P = 0.046$	$r = -0.375$ $P = 0.022$
BMI	$Rho = -0.228$ $P = 0.175$	$r = -0.225$ $P = 0.180$
FEV1 (% pred)	$Rho = -0.316$ $P = 0.057$	$r = -0.144$ $P = 0.395$
FVC (% pred)	$Rho = 0.025$ $P = 0.885$	$r = -0.279$ $P = 0.094$
FEV1/FVC	$Rho = -0.308$ $P = 0.064$	$r = 0.024$ $P = 0.886$

breakdown of antioxidants in favor of the reduction thereof occurs oxidative stress^{22,23}. Oxidative stress not only damages lungs but activates molecular mechanisms performing lung inflammation, as well²⁴. Increased oxidative stress parameters and decreased BMI values in our study are parallel to these studies. MDA levels of COPD patient groups [1st, 15th day and total) were significantly higher than the controls in our study. This increase was even more apparent in patients with COPD exacerbation. This shows that oxidant effect increasing in COPD leads to lipid peroxidation. Similarly, MDA levels were statistically significantly high in COPD patients^{25,26}. Likewise, in a lot of literature studies, MDA levels were higher in acute exacerbation period compared to the stable period in COPD patients similar to the data of our study. Again in these studies, the decreases detected in antioxidant enzyme levels indicate that oxidant/antioxidant balance is break down in favor of oxidants in COPD patients^{27,28}.

It is detected that in studies that are made considering the fact that pathology is not only limited to airways in COPD, but there are also the systemic effects, weight loss is associated with inflammatory cytokines such as serum IL-6 and TNF- α and adipocytokines such as serum leptin, adiponectin and

visfatin^{29,30}. It is reported that adiponectin, which is among the adipokines regulating the energy metabolism and appetite, increases insulin sensitivity and the levels decrease in obese individuals as inflammatory response³¹. Anti-inflammatory effect of adiponectin manifests itself as the increase in plasma adiponectin levels in COPD individuals³². Carolan *et al.* also detected that increased plasma adiponectin levels in COPD are associated with lung emphysema and low bronchial capacity related to BMI decreasing at later ages leads to disease progression³². In our study, similarly, serum adiponectin levels were significantly higher both in COPD exacerbation and stable COPD groups compared to the controls. Significant increase of adiponectin levels in COPD groups with low BMI is parallel to the mechanisms stated above. These results give rise to the thought that hyperinflation and low BMI in COPD patients increase serum adiponectin levels with anti-inflammatory effect and contribute to the defense.

Intra-abdominal adipose tissue, an active endocrine organ, has a positive correlation with pro-inflammatory adipocytokines (IL-6, TNF- α and leptin) regulating insulin sensitivity and negative correlation with adiponectin³³. When (1st, 15th day and total) serum IL-6 levels of COPD patient group were compared with the control group, there was a statistically significant decrease, and there was a negative correlation between decreased IL-6 levels and serum adiponectin levels ($r = -0.375$, $P = 0.22$). Except for the increase in acute phase proteins, functional effects of IL-6 in circulation has not been determined yet, but there is evidence regarding that it may be associated with skeletal muscle weakness³⁴. It is shown that there is a correlation between the increased IL-6 concentrations in circulation and systemic inflammation degree in some comorbid diseases. Inflammatory mediators such as TNF- α , IL-1 β and IL-6 can be accepted as general markers of inflammation. However, IL-6 is not a simple inflammatory marker and has an active role in the pathogenesis of some diseases such as rheumatoid arthritis³⁵. Studies claiming that IL-6 has a potential role in the pathogenesis in COPD and induced mucus IL-6 levels are inversely proportional to FEV1³⁶.

In addition to the studies stating that visfatin levels in circulation system of the patients losing excessive weight after the stomach surgery³⁷. Both high³⁸ and low³⁹ visfatin levels were reported in COPD patient groups when compared to the control group. In our

study, serum visfatin levels of COPD patient group (1st, 15th day and total) were statistically significantly lower when compared to the control group. Visfatin, along with TNF- α and IL-6, plays a key role in the regulation of inflammatory process⁴⁰. COPD patient data studying on visfatin level are quite contradictory. In addition to the studies reporting high visfatin levels in COPD patients with low BMI^{40,41}, there are also studies detecting decreased visfatin levels^{14,42}. In a study, it was reported that there is a positive correlation between low plasma visfatin levels and visfatin and IL-6 in COPD patients¹⁴. Similarly, in our study, there was a positive correlation between visfatin and IL-6 levels in COPD exacerbation patient group ($r = 0.330$, $P = 0.046$).

Conclusion

It was suggested that the level changes of MDA, IL-6, visfatin and adiponectin may be an important factor in the development of COPD. Meanwhile, the studies in which the correlation between systemic inflammation and weight loss during COPD is evaluated, although a lot of inflammatory markers are correlated with weight loss and other systems, the mechanisms could not be identified clearly. Methods should be developed by considering the effects of decreased BMI on the disease severity in the fight against COPD. Moreover, while treatment protocols for the increased inflammation and oxidative stress are developed, thanks to the assessments to be made in cytokine and protein changes with pro-inflammatory/anti-inflammatory effect by considering the other systems accompanying the disease in addition to the respiratory system, early diagnosis and the treatment of the accompanying diseases, COPD morbidity and mortality can be reduced.

Acknowledgement

The authors acknowledge the contributions from medical biochemistry research assistants who helped ELISA studies.

Conflict of interest

All authors declare no conflict of interest.

References

- 1 Sepúlveda-Loyola W, Osadnik C, Phu S, Morita AA, Duque G & Probst VS, Diagnosis, prevalence, and clinical impact of sarcopenia in copd: A systematic review and meta-analysis. *J Cachexia Sarcopenia Muscle*, 11 (2020) 1164.
- 2 Capron T, Bourdin A, Perez T & Chanez P, COPD beyond proximal bronchial obstruction: phenotyping and related tools at the bedside. *Eur Respir Rev*, 28 (2019) 1.
- 3 Cavaillès A, Brinchault-Rabin G, Dixmier A, Goupil F, Gut-Gobert C, Marchand-Adam S, Meurice JC, Morel H,

- Person-Tacnet C, Leroyer C & Diot P, Comorbidities of COPD. *Eur Respir Rev*, 22 (2013) 454.
- 4 Liu G & Summer R, Cellular metabolism in lung health and disease. *Annu Rev Physiol*, 81 (2019) 403.
 - 5 Ritchie AI & Wedzicha JA, Definition, causes, pathogenesis, and consequences of chronic obstructive pulmonary disease exacerbations. *Clin Chest Med*, 41 (2020) 421.
 - 6 Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, Gargiulo G, Testa G, Cacciatore F, Bonaduce D & Abete P, Oxidative stress, aging, and diseases. *Clin Interv Aging*, 13 (2018) 757.
 - 7 Choi HM, Doss HM & Kim KS, Multifaceted physiological roles of adiponectin in inflammation and diseases. *Int J Mol Sci*, 21 (2020) 1219.
 - 8 Ramanathan V, Raj DA & Thekkumalai M, *Macrotyloma unijflorum* (Lam.) Verdc. improves glucose metabolism and proinflammatory parameters in high fructose fed rats. *Indian J Exp Biol*, 57 (2019) 594.
 - 9 Lee YH & Bae SC, Circulating adiponectin and visfatin levels in rheumatoid arthritis and their correlation with disease activity: A meta-analysis. *Int J Rheum Dis*, 21 (2018) 664.
 - 10 Karmiris K, Koutroubakis IE, Xidakis C, Polychronaki M, Voudouri T & Kouroumalis EA, Circulating levels of leptin, adiponectin, resistin, and ghrelin in inflammatory bowel disease. *Inflamm Bowel Dis*, 12 (2006) 100.
 - 11 Ilhan N, Susam S, Canpolat O & Belhan O, The emerging role of leptin, adiponectin and visfatin in ischemic/hemorrhagic stroke. *Br J Neurosurg*, 33 (2019) 504.
 - 12 Lindberg S, Jensen JS, Bjerre M, Frystyk J, Flyvbjerg A, Jeppesen J & Mogelvang R, Low adiponectin levels at baseline and decreasing adiponectin levels over 10 years of follow-up predict risk of the metabolic syndrome. *Diabetes Metab*, 43 (2017) 134.
 - 13 Rajesh Y & Sarkar D, Association of adipose tissue and adipokines with development of obesity-induced liver cancer. *Int J Mol Sci*, 22 (2021) 2163.
 - 14 Leivo-Korpela S, Lehtimäki L, Hämäläinen M, Vuolteenaho K, Kööbi L, Järvenpää R, Kankaanranta H, Saarelainen S & Moilanen E, Adipokines NUCB2/nesfatin-1 and visfatin as novel inflammatory factors in chronic obstructive pulmonary disease. *Mediators Inflamm*, 2014 (2014) 1.
 - 15 Rincon M & Irvin CG, Role of IL-6 in asthma and other inflammatory pulmonary diseases. *Int J Biol Sci*, 8 (2012) 1281.
 - 16 Liang R, Zhang W & Song YM, Levels of leptin and IL-6 in lungs and blood are associated with the severity of chronic obstructive pulmonary disease in patients and rat models. *Mol Med Rep*, 7 (2013) 1470.
 - 17 Rutten EP, Calverley PM, Casaburi R, Agusti A, Bakke P, Celli B, Coxson HO, Crim C, Lomas DA, Macnee W, Miller BE, Rennard SI, Scanlon PD, Silverman EK, Tal-Singer R, Vestbo J, Watkins ML & Wouters EF, Changes in body composition in patients with chronic obstructive pulmonary disease: do they influence patient-related outcomes? *Ann Nutr Metab*, 63 (2013) 239.
 - 18 Chen W, Sadatsafavi M, FitzGerald JM, Lynd LD & Sin DD, Gender modifies the effect of body mass index on lung function decline in mild-to-moderate COPD patients: a pooled analysis. *Respir Res*, 22 (2021) 1.
 - 19 Scoditti E, Massaro M, Garbarino S & Toraldo DM, Role of diet in chronic obstructive pulmonary disease prevention and treatment. *Nutrients*, 11 (2019) 1357.
 - 20 Kwan HY, Maddocks M, Nolan CM, Jones SE, Patel S, Barker RE, Kon SSC, Polkey MI, Cullinan P & Man WD, The prognostic significance of weight loss in chronic obstructive pulmonary disease-related cachexia: a prospective cohort study. *J Cachexia Sarcopenia Muscle*, 10 (2019) 1330.
 - 21 Domej W, Oettl K & Renner W, Oxidative stress and free radicals in COPD--implications and relevance for treatment. *Int J Chron Obstruct Pulmon Dis*, 9 (2014) 1207.
 - 22 Mistry KN, Dabhi BK & Joshi BB, Evaluation of oxidative stress biomarkers and inflammation in pathogenesis of diabetes and diabetic nephropathy. *Indian J Biochem Biophys*, 57 (2020) 45.
 - 23 Aktas HS, Ozel Y, Ahmad S, Pençe HH, Sayir N, Sapmaz T, Özçelik F & Elçioglu HK. Influence of walnut on hepatic ischemia-reperfusion injury in streptozotocin-induced diabetic rats. *Indian J Biochem Biophys*, 58 (2021) 45.
 - 24 Pizzino G, Irrera N, Cucinotta M, Pallio G, Mannino F, Arcoraci V, Squadrito F, Altavilla D & Bitto A, Oxidative stress: Harms and benefits for human health. *Oxid Med Cell Longev*, 2017 (2017) 1.
 - 25 Pandey S, Garg R, Kant S & Gaur P, Vitamin D, C-reactive protein, and oxidative stress markers in chronic obstructive pulmonary disease. *Tzu Chi Med J*, 33 (2021) 80.
 - 26 Wang J, Li Y, Zhao P, Tian Y, Liu X, He H, Jia R, Oliver BG & Li J, Exposure to air pollution exacerbates inflammation in rats with preexisting COPD. *Mediators Inflamm*, 2020 (2020).
 - 27 Antus B, Harnasi G, Drozdovszky O & Barta I, Monitoring oxidative stress during chronic obstructive pulmonary disease exacerbations using malondialdehyde. *Respirology*, 19 (2014) 74.
 - 28 Stanojkovic I, Kotur-Stevuljjevic J, Milenkovic B, Spasic S, Vujic T, Stefanovic A, Llic A & Ivanisevic J, Pulmonary function, oxidative stress and inflammatory markers in severe COPD exacerbation. *Respir Med*, 105 (2011) 31.
 - 29 Wang Y, Xu J, Meng Y, Adcock IM & Yao X, Role of inflammatory cells in airway remodeling in COPD. *Int J Chron Obstruct Pulmon Dis*, 13 (2018) 3341.
 - 30 Chwalba A, Machura E, Ziora K & Ziora D, The role of adipokines in the pathogenesis and course of selected respiratory diseases. *Endokrynol Pol*, 70 (2019) 504.
 - 31 Knights AJ, Funnell AP, Pearson RC, Crossley M & Bell-Anderson KS, Adipokines and insulin action: A sensitive issue. *Adipocyte*, 3 (2014) 88.
 - 32 Carolan BJ, Kim YI, Williams AA, Kechris K, Lutz S, Reisdorph N & Bowler RP, The association of adiponectin with computed tomography phenotypes in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*, 188 (2013) 561.
 - 33 Jung UJ & Choi MS, Obesity and its metabolic complications: the role of adipokines and the relationship between obesity, inflammation, insulin resistance, dyslipidemia and nonalcoholic fatty liver disease. *Int J Mol Sci*, 15 (2014) 6184.
 - 34 Yende S, Waterer GW, Tolley EA, Newman AB, Bauer DC, Taaffe DR, Jensen R, Crapo R, Rubin S, Nevitt M, Simonsick EM, Satterfield S, Harris T & Kritchevsky SB, Inflammatory markers are associated with ventilatory limitation and muscle dysfunction in obstructive lung disease in well-functioning elderly subjects. *Thorax*, 61 (2006) 10.
 - 35 Kishimoto T, IL-6: from its discovery to clinical applications. *Int Immunol*, 22 (2010) 347.

- 36 Grubek-Jaworska H, Paplińska M, Hermanowicz-Salamon J, Białek-Gosk K, Dąbrowska M, Grabczak E, Domagała-Kulawik J, Stępień J & Chazan R, IL-6 and IL-13 in induced sputum of COPD and asthma patients: correlation with respiratory tests. *Respiration*, 84 (2012) 101.
- 37 Hosseinzadeh-Attar MJ, Golpaie A, Janani L & Derakhshanian H, Effect of weight reduction following bariatric surgery on serum visfatin and adiponectin levels in morbidly obese subjects. *Obes Facts*, 6 (2013) 193.
- 38 Pérez-Bautista O, Montaña M, Pérez-Padilla R, Zúñiga-Ramos J, Camacho-Priego M, Barrientos-Gutiérrez T, Buendía-Roldan I, Velasco-Torres Y & Ramos C, Women with COPD by biomass show different serum profile of adipokines, incretins, and peptide hormones than smokers. *Respir Res*, 19 (2018) 1.
- 39 Göktepe M, Korkmaz C, Zamani A, Demirbaş S & Kılınc İ, Evaluation of serum resistin, visfatin, and chemerin levels in patients with lung cancer and chronic obstructive pulmonary disease. *Turk Thorac J*, 21 (2020) 169.
- 40 Makki K, Froguel P & Wolowczuk I, Adipose tissue in obesity-related inflammation and insulin resistance: cells, cytokines, and chemokines. *ISRN Inflamm*, 2013 (2013) 1.
- 41 Moschen AR, Kaser A, Enrich B, Mosheimer B, Theurl M, Niederegger H & Tilg H, Visfatin, an adipocytokine with proinflammatory and immunomodulating properties. *J Immunol*, 178 (2007) 1748.
- 42 Eker S, Ayaz L, Tamer L & Ulubas B, Leptin, visfatin, insulin resistance, and body composition change in chronic obstructive pulmonary disease. *Scand J Clin Lab Invest*, 70 (2010) 40.