



Free radical scavenging and α -glucosidase inhibitory activity of (*E*)-methyl/ethyl-3-(2-hydroxyphenyl)acrylates

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(*E*)-Methyl/ethyl-3-(2-hydroxyphenyl)acrylates **3a-x** have been prepared by the reaction of salicylaldehydes **1a-l** with Wittig reagents such as methyl (triphenylphosphoranylidene)acetate **2a** and ethyl (triphenylphosphoranylidene)acetate **2b** in dry DCM at room temperature. All the synthesized compounds have been evaluated for free-radical scavenging and α -glucosidase inhibitory activities. Compounds **3c** and **3d** display DPPH free radical scavenging activity. All the compounds have shown ABTS free radical scavenging activity except four compounds **3s-t** and **3w-x**. Compounds **3g**, **3p** and **3r** display α -glucosidase inhibitory activity.

Keywords: (*E*)-Methyl/ethyl-3-(2-hydroxyphenyl)acrylates, salicylaldehydes, free radical scavenging, α -glucosidase inhibitory activity

(*E*)- α,β -Unsaturated esters¹ are an important compounds and valuable intermediates for the preparation of various synthetic and natural products². The widely used methods for the preparation of these esters are the Wittig reaction³ and Horner-Wadsworth-Emmons using alkoxy carbonyl methylene(triphenyl)phosphoranes and trialkyl phosphonoacetates³⁻¹⁰. These esters have applications in the food, polymer and perfume industries. Therefore, the commercial scale preparations of these compounds are necessary.

As part of our ongoing research on the preparation of heterocyclic compounds^{11,12} and natural products^{13,14}, recently, we have prepared various novel heterocyclic compounds by the reaction of salicylaldehydes with β -ketoesters¹⁵⁻¹⁸. The reaction of salicylaldehydes with Wittig reagent provides the (*E*)- α,β -unsaturated esters and its biological properties have not been studied. Therefore, the present manuscript describes the preparation of α,β -unsaturated esters and their evaluation of DPPH, ABTS⁺ free radical scavenging and α -glucosidase inhibitory activities.

Results and Discussion

The target (*E*)-methyl/ethyl 3-(2-hydroxyphenyl)acrylate compounds **3a-x** have been

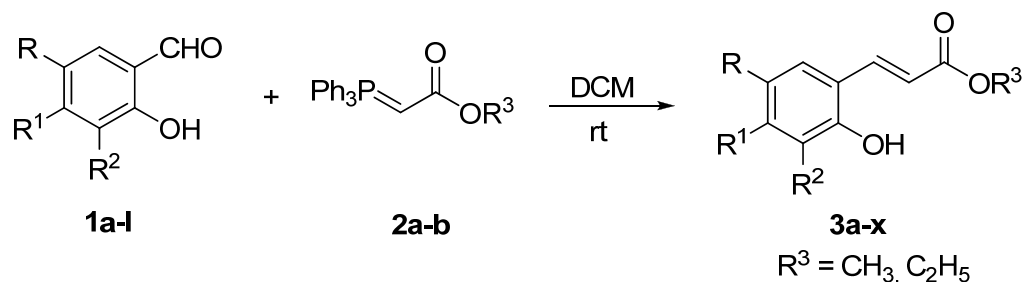
prepared by the reaction of salicylaldehydes **1a-l** with Wittig reagents¹⁹ namely methyl (triphenylphosphoranylidene)acetate **2a** and ethyl (triphenylphosphoranylidene)acetate **2b** in dry DCM at RT (Scheme I, Table I). All the compounds are characterized by spectral data.

Biology

The prepared compounds **3a-x** have been evaluated for their biological activities such as DPPH, ABTS⁺ free radical scavenging and α -glucosidase inhibitory²⁰ for the first time and the results are described below.

Free radicals scavenging activity

The DPPH and ABTS⁺ free radical scavenging activity of compounds **3a-x** are presented in Table II along with the standard drugs Ascorbic acid and Trolox (SC₅₀ values)²⁰. Among the prepared compounds, the methoxy (5th position) substituted compounds **3c** (SC₅₀ 6.47 μ g/mL) and **3d** (SC₅₀ 6.21 μ g/mL) have shown the DPPH free radical scavenging activity in comparison with the standard compound Ascorbic acid (SC₅₀ 4.08 μ g/mL). However, the methoxy compounds **3e-f** positioned at 4th position and remaining compounds **3a-b** and **3g-x** could not display the activity.



Scheme I

Table I — Preparation of (*E*)-methyl/ethyl-3-(2-hydroxyphenyl) acrylates **3a-x**

Entry	Compd	R	R ¹	R ²	R ³	Yield ^a (%)
1	3a	H	H	H	CH ₃	84
2	3b	H	H	H	C ₂ H ₅	86
3	3c	OCH ₃	H	H	CH ₃	82
4	3d	OCH ₃	H	H	C ₂ H ₅	84
5	3e	H	OCH ₃	H	CH ₃	82
6	3f	H	OCH ₃	H	C ₂ H ₅	82
7	3g	CH ₃	H	H	CH ₃	76
8	3h	CH ₃	H	H	C ₂ H ₅	80
9	3i	F	H	H	CH ₃	70
10	3j	F	H	H	C ₂ H ₅	72
11	3k	Cl	H	H	CH ₃	74
12	3l	Cl	H	H	C ₂ H ₅	75
13	3m	Br	H	H	CH ₃	77
14	3n	Br	H	H	C ₂ H ₅	79
15	3o	Cl	H	Cl	CH ₃	74
16	3p	Cl	H	Cl	C ₂ H ₅	74
17	3q	Br	H	Br	CH ₃	68
18	3r	Br	H	Br	C ₂ H ₅	70
19	3s	NO ₂	H	H	CH ₃	62
20	3t	NO ₂	H	H	C ₂ H ₅	65
21	3u	NO ₂	H	Br	CH ₃	56
22	3v	NO ₂	H	Br	C ₂ H ₅	54
23	3w	NO ₂	H	NO ₂	CH ₃	48
24	3x	NO ₂	H	NO ₂	C ₂ H ₅	42

^a Isolated yields

The ABTS⁺ free radical scavenging activity of compounds **3a-r** and **3u-v** have shown ranging from SC₅₀ 1.24-3.93 μg/mL when compared to standard compound Trolox (SC₅₀ 1.16 μg/mL). It is interesting to note that twenty compounds (methyl, methoxy, halo substitution) have shown ABTS⁺ free radical scavenging activity among the prepared 24 compounds. The nitro substituted compounds **3s-t** and di nitro substituted compounds **3w-x** could not shown ABTS⁺ free radical scavenging activity.

α-Glucosidase inhibitory activity

α-Glucosidase inhibitory activity of compounds **3a-x** and their IC₅₀ values presented in Table II along with the standard drug Acarbose²⁰. Three compounds have

shown α-glucosidase inhibitory activity in the present series of compounds. The methyl substituted compound **3g** (IC₅₀ 4.39 μg/mL) has shown better α-glucosidase inhibitory activity when compared to methoxy **3c-f**, and halo **3i-o**. Interestingly, the dichloro **3p** (IC₅₀ 10.79 μg/mL) shown moderate activity and dibromo compound **3r** (IC₅₀ 2.31 μg/mL) displayed potent α-glucosidase inhibitory activity (Table II).

Experimental Section

All the chemicals and reagents were purchased from Aldrich (Sigma-Aldrich, USA), AVRA Chemicals Pvt. Ltd (Hyderabad, India) and were used without further purification. Reactions were monitored by thin layer chromatography (TLC) on pre-coated silica gel 60 F₂₅₄ (mesh); spots were visualized under UV light. Melting points were determined on a Stuart melting point apparatus and are uncorrected. IR spectrum was recorded with a Thermo Nicolet Nexus 670 FT spectrometer. ¹H and ¹³C NMR spectra were recorded on Bruker Avance 300, 400 and 500 MHz spectrometers. Chemical shifts (δ) are quoted in parts per million and are referenced to tetramethylsilane (TMS) as internal standard. ESI-MS were obtained on quato micro spectrometer.

General experimental procedure for the preparation of compounds, 3a-x

Methyl (triphenylphosphoranylidene)acetate **2a** (1.0 mmol) was added to a stirred solution of salicylaldehyde **1a-l** (1.0 mmol) in dry DCM (2 mL) at RT. The reaction mixture was stirred at the same temperature until the starting materials disappeared (TLC). After completion of the reaction, the solvent was removed and the reaction mixture was purified by column chromatography (EtOAc/hexane) to give colourless solid **3a**. Similarly, the compounds **3b-x** have been prepared by the reaction of salicylaldehydes **1a-l** with Wittig reagents such as methyl (triphenylphosphoranylidene)acetate **2a** and

Table II — DPPH, ABTS⁺, α -glucosidase inhibitory activity profile of compounds **3a-x**

Compd	DPPH % Inhibition	ABTS ⁺ % Inhibition	AGI % Inhibition
	25 μ g/mL (SC ₅₀ μ g /mL)	20 μ g /mL (SC ₅₀ μ g /mL)	20 μ g/mL (IC ₅₀ μ g /mL)
3a	12.00 \pm 0.07	98.19 \pm 0.32 (1.25)	25.79 \pm 0.62
3b	10.42 \pm 0.22	96.71 \pm 0.48 (1.34)	35.12 \pm 0.61
3c	72.75 \pm 0.04 (6.47)	96.37 \pm 0.64 (1.36)	ND
3d	73.17 \pm 1.23 (6.21)	98.19 \pm 0.00 (1.25)	19.72 \pm 0.00
3e	40.41 \pm 0.86	97.28 \pm 0.00 (1.41)	19.01 \pm 0.00
3f	42.07 \pm 0.07	97.28 \pm 0.00 (1.31)	19.01 \pm 0.00
3g	26.56 \pm 1.49	98.64 \pm 0.00 (1.24)	80.19 \pm 1.87(4.39)
3h	26.48 \pm 0.26	96.37 \pm 1.28 (1.36)	32.31 \pm 0.87
3i	14.43 \pm 0.45	97.51 \pm 0.96 (1.37)	34.33 \pm 2.49
3j	12.08 \pm 0.93	97.96 \pm 0.00 (1.29)	29.58 \pm 0.75
3k	6.86 \pm 0.04	97.28 \pm 0.00 (1.31)	38.38 \pm 0.50
3l	6.30 \pm 0.45	97.51 \pm 0.64 (1.37)	40.41 \pm 0.12
3m	5.20 \pm 0.75	97.51 \pm 0.00 (1.37)	42.96 \pm 0.00
3n	4.35 \pm 0.30	97.51 \pm 0.32 (1.37)	34.15 \pm 0.25
3o	2.27 \pm 1.21	98.19 \pm 0.00 (1.25)	57.31 \pm 1.62
3p	3.38 \pm 0.48	98.41 \pm 0.00 (1.38)	64.96 \pm 0.00 (10.79)
3q	ND	71.32 \pm 1.12 (3.93)	52.90 \pm 1.89
3r	3.35 \pm 0.37	96.71 \pm 0.16 (1.34)	97.36 \pm 0.24 (2.31)
3s	ND	5.22 \pm 1.28	53.70 \pm 0.25
3t	ND	3.29 \pm 0.16	ND
3u	8.52 \pm 1.72	98.64 \pm 0.00 (1.24)	28.61 \pm 0.37
3v	8.47 \pm 1.198	98.53 \pm 0.16 (1.26)	38.91 \pm 0.00
3w	ND	6.24 \pm 0.16	37.68 \pm 0.00
3x	ND	2.72 \pm 0.00	ND
Ascorbic acid	85.68 \pm 0.86 (4.08)	—	—
Trolox	—	98.87 \pm 0.00 (1.16)	—
Acarbose	—	—	98.57 \pm 0.05 (2.17)

ethyl (triphenylphosphoranylidene)acetate **2b** under our optimized conditions. All the compounds are characterized by spectral data.

Methyl(*E*)-3-(2-hydroxyphenyl)arylate, 3a: Colourless solid. m.p.136-137°C. ¹H NMR (500 MHz, CDCl₃): δ 8.06 (d, *J* = 16.2 Hz, 1H, CH), 7.47 (dd, *J* = 7.7, 1.2 Hz, 1H, aromatic), 7.27-7.21 (m, 1H, aromatic), 6.92 (t, *J* = 7.5 Hz, 1H, aromatic), 6.86 (dd, *J* = 8.1, 0.7 Hz, 1H, aromatic), 6.72 (s, 1H, OH), 6.65 (d, *J* = 16.2 Hz, 1H, CH), 3.83 (s, 3H, OCH₃); ESI-MS: *m/z* [M+H]⁺ 179.

Ethyl(*E*)-3-(2-hydroxyphenyl)acrylate, 3b: Colourless solid. m.p.143-144°C. ¹H NMR (400 MHz, CDCl₃): δ 8.08 (d, *J* = 16.1 Hz, 1H, CH), 7.45 (s, 2H, aromatic), 7.22 (t, *J* = 7.2 Hz, 1H, aromatic), 6.89 (d, *J* = 6.9 Hz, 2H, aromatic, OH), 6.67 (d, *J* = 16.1 Hz, 1H, CH), 4.30 (d, *J* = 6.9 Hz, 2H, OCH₂), 1.33 (dd, *J* = 24.8, 18.2 Hz, 3H, OCH₃); ESI-MS: *m/z* [M+H]⁺ 193.

Methyl(*E*)-3-(2-hydroxy-5-methoxyphenyl)-acrylate, 3c: Colourless solid. m.p.126-130°C. ¹H NMR (400 MHz, CDCl₃): δ 8.03 (d, *J* = 16.2 Hz, 1H, CH), 6.97 (d, *J* = 2.6 Hz, 1H, aromatic), 6.86-6.77 (m, 2H, aromatic), 6.62 (s, 1H, OH), 6.58 (d, *J* =

16.1 Hz, 1H, CH), 3.83 (d, *J* = 5.4 Hz, 3H, OCH₃), 3.77 (s, 3H, OCH₃); ESI-MS: *m/z* [M+H]⁺ 209.

Ethyl(*E*)-3-(2-hydroxy-5-methoxyphenyl)-acrylate, 3d: Colourless solid. m.p.129-134°C. ¹H NMR (400 MHz, CDCl₃): δ 8.02 (d, *J* = 16.1 Hz, 1H, CH), 6.98 (d, *J* = 2.8 Hz, 1H, aromatic), 6.82 (m, 2H, aromatic), 6.57 (d, *J* = 16.1 Hz, 1H, aromatic), 6.31 (s, 1H, OH), 4.35-4.24 (m, 2H, OCH₂), 3.83-3.73 (m, 3H, OCH₃), 1.40-1.31 (m, 3H, OCH₃); ESI-MS: *m/z* [M+H]⁺ 223.

Methyl(*E*)-3-(2-hydroxy-4-methoxyphenyl)-acrylate, 3e: Colourless solid. m.p.126-128°C. ¹H NMR (400 MHz, CDCl₃): δ 7.98 (d, *J* = 16.1 Hz, 1H, CH), 7.39 (d, *J* = 8.7 Hz, 1H, aromatic), 7.04 (s, 1H, OH), 6.52 (d, *J* = 16.1 Hz, 1H, CH), 6.49 (dd, *J* = 8.7, 2.4 Hz, 1H, aromatic), 6.41 (d, *J* = 2.4 Hz, 1H, aromatic), 3.83 (s, 3H, OCH₃), 3.80 (s, 3H, OCH₃); ESI-MASS: *m/z* [M+H]⁺ 209.

Ethyl(*E*)-3-(2-hydroxy-4-methoxyphenyl)-acrylate, 3f: Colourless solid. m.p.129-134°C. ¹H NMR (500 MHz, CDCl₃): δ 8.00 (d, *J* = 16.1 Hz, 1H, CH), 7.39 (d, *J* = 8.7 Hz, 1H, aromatic), 7.32 (s, 1H, OH), 6.54 (d, *J* = 16.1 Hz, 1H, CH), 6.48 (dd, *J* = 8.6, 2.3 Hz, 1H, aromatic), 6.42 (d, *J* = 2.4 Hz, 1H,

aromatic), 4.33-4.24 (m, 2H, OCH₂), 3.79 (s, 3H, OCH₃), 1.39-1.29 (m, 3H, Θ CH₃); ESI-MS: m/z [M+H]⁺ 223.

Methyl(*E*)-3-(2-hydroxy-5-methylphenyl)-acrylate, 3g: Colourless solid. m.p.136-144°C. ¹H NMR (400 MHz, CDCl₃): δ 8.01 (d, J = 16.2 Hz, 1H, CH), 7.04 (dd, J = 8.2, 1.7 Hz, 1H, aromatic), 6.75 (d, J = 8.2 Hz, 1H, aromatic), 6.61 (d, J = 16.2 Hz, 1H, CH), 6.38 (s, 1H, OH), 3.88-3.77 (m, 3H, OCH₃), 2.27 (s, 3H, CH₃); ESI-MS: m/z [M+H]⁺ 193.

Ethyl(*E*)-3-(2-hydroxy-5-methylphenyl)-acrylate, 3h: Colourless solid. m.p.132-134°C. ¹H NMR (500 MHz, CDCl₃): δ 7.97 (d, J = 16.1 Hz, 1H, CH), 7.26 (s, 1H, aromatic), 7.03 (dd, J = 8.2, 1.7 Hz, 1H, aromatic), 6.75 (t, J = 6.1 Hz, 1H), 6.59 (dd, J = 16.1, 5.4 Hz, 1H, CH), 6.24 (s, 1H, OH), 4.32-4.24 (m, 2H, OCH₂), 2.27 (s, 3H, CH₃), 1.34 (t, J = 7.1 Hz, 3H, Θ CH₃); ESI-MS: m/z [M+H]⁺ 207.

Methyl(*E*)-3-(5-fluoro-2-hydroxyphenyl)-acrylate, 3i: Colourless solid. m.p.126-128°C. ¹H NMR (500 MHz, CDCl₃): δ 7.97 (d, J = 16.2 Hz, 1H, CH), 7.16 (dd, J = 9.1, 3.0 Hz, 1H, aromatic), 6.95 (m, Hz, 1H, aromatic), 6.80 (dd, J = 8.9, 4.5 Hz, 1H, aromatic), 6.56 (d, J = 16.2 Hz, 1H, CH), 6.22 (d, J = 23.5 Hz, 1H, OH), 3.83 (d, J = 3.4 Hz, 3H, CH₃). ESI-MS: m/z [M+H]⁺ 197.

Ethyl(*E*)-3-(5-fluoro-2-hydroxyphenyl)acrylate, 3j: Colourless solid. m.p.125-126°C. ¹H NMR (400 MHz, CDCl₃): δ 8.02 (d, J = 16.2 Hz, 1H, CH), 7.16 (dd, J = 9.2, 3.0 Hz, 1H, aromatic), 6.95 (ddd, J = 8.8, 7.8, 3.0 Hz, 1H, aromatic), 6.85-6.78 (m, 1H, aromatic), 6.59-6.54 (d, J = 16.2 Hz, 1H, CH), 4.29 (q, J = 7.1 Hz, 2H, OCH₂), 1.39-1.32 (m, 3H, Θ CH₃); ESI-MS: m/z [M+H]⁺ 211.

Methyl(*E*)-3-(5-chloro-2-hydroxyphenyl)-acrylate, 3k: Colourless solid. m.p.134-136°C. ¹H NMR (400 MHz, CDCl₃): δ 7.92 (d, J = 16.2 Hz, 1H, CH), 7.44 (d, J = 2.5 Hz, 1H, aromatic), 7.19 (dd, J = 8.6, 2.5 Hz, 1H, aromatic), 6.79 (d, J = 8.6 Hz, 1H, aromatic), 6.58 (d, J = 16.2 Hz, 1H, CH), 6.30 (s, 1H, OH), 3.83 (s, 3H, OCH₃); ESI-MS: m/z [M+H]⁺ 213.

Ethyl(*E*)-3-(5-chloro-2-hydroxyphenyl)acrylate, 3l: Colourless solid. m.p.124-126°C. ¹H NMR (400 MHz, CDCl₃): δ 8.00 (d, J = 16.2 Hz, 1H, CH), 7.72 (s, 1H, OH), 7.42 (d, J = 2.5 Hz, 1H, aromatic), 7.17 (dd, J = 8.6, 2.5 Hz, 1H, aromatic), 6.83 (d, J = 8.6 Hz, 1H, aromatic), 6.64 (d, J = 16.2 Hz, 1H, CH), 4.30 (q, J = 7.1 Hz, 2H, OCH₂), 1.36 (t, J = 7.1 Hz, 3H, Θ CH₃); ESI-MS: m/z [M+H]⁺ 227.

Methyl(*E*)-3-(5-bromo-2-hydroxyphenyl)-acrylate, 3m: Colourless solid. m.p. 135-137°C. ¹H NMR (400 MHz, CDCl₃): δ 7.93 (d, J = 16.2 Hz, 1H, CH), 7.58 (d, J = 2.3 Hz, 1H, aromatic), 7.32 (dd, J = 8.6, 2.4 Hz, 1H, aromatic, aromatic), 6.75 (d, J = 8.6 Hz, 1H, aromatic), 6.72 (s, 1H, OH), 6.59 (d, J = 16.2 Hz, 1H, CH), 3.83 (s, 3H, OCH₃); ESI-MS: m/z [M+H]⁺ 257.

Ethyl(*E*)-3-(5-bromo-2-hydroxyphenyl)-acrylate, 3n: Colourless solid. m.p.138-140°C. ¹H NMR (400 MHz, CDCl₃): 7.97 (d, J = 16.2 Hz, 1H, CH), 7.57 (d, J = 2.4 Hz, 1H, aromatic), 7.51 (s, 1H, OH), 7.36-7.27 (m, 1H, aromatic), 6.78 (d, J = 8.6 Hz, 1H, aromatic), 6.63 (d, J = 16.2 Hz, 1H, CH), 4.37-4.25 (m, 2H, OCH₂), 1.41-1.30 (m, 3H, Θ CH₃); ESI-MS: m/z [M+H]⁺ 271.

Methyl(*E*)-3-(3,5-dichloro-2-hydroxyphenyl)-acrylate, 3o: Colourless solid. m.p.128-130°C. ¹H NMR (400 MHz, CDCl₃): δ 7.86 (d, J = 16.2 Hz, 1H, CH), 7.37 (dd, J = 11.6, 2.4 Hz, 2H, aromatic, CH), 6.57 (d, J = 16.2 Hz, 1H, CH), 6.08 (s, 1H, OH), 3.82 (s, 3H, OCH₃); ESI-MS: m/z [M+H]⁺ 247.

Ethyl(*E*)-3-(3,5-dichloro-2-hydroxyphenyl)-acrylate, 3p: Colourless solid. m.p.119-120°C. ¹H NMR (500 MHz, CDCl₃): δ 7.85 (d, J = 16.2 Hz, 1H, CH), 7.37 (dd, J = 18.8, 2.4 Hz, 2H, aromatic, CH), 6.57 (d, J = 16.2 Hz, 1H, CH), 6.04 (s, 1H, OH), 4.30-4.25 (m, 2H, OCH₂), 1.37-1.32 (m, 3H, Θ CH₃); ESI-MS: m/z [M+H]⁺ 261.

Methyl(*E*)-3-(3,5-dibromo-2-hydroxyphenyl)-acrylate, 3q: Colourless solid. m.p.122-124°C. ¹H NMR (500 MHz, CDCl₃): δ 7.84 (d, J = 16.2 Hz, 1H, CH), 7.61 (d, J = 2.3 Hz, 1H, aromatic), 7.55 (d, J = 2.3 Hz, 1H, aromatic), 6.56 (d, J = 16.1 Hz, 1H, CH), 5.97 (s, 1H, OH), 3.81 (s, 3H, OCH₃). ESI-MS: m/z [M+H]⁺ 335.

Ethyl(*E*)-3-(3,5-dibromo-2-hydroxyphenyl)-acrylate, 3r: Colourless solid. m.p. 125-127°C. ¹H NMR (500 MHz, CDCl₃): δ 7.84 (d, J = 16.2 Hz, 1H, CH), 7.60 (d, J = 2.3 Hz, 1H, aromatic), 7.56 (d, J = 2.3 Hz, 1H, aromatic), 6.55 (d, J = 16.1 Hz, 1H, CH), 6.04 (s, 1H, OH), 4.27 (q, J = 7.1 Hz, 2H, OCH₂), 1.34 (dd, J = 9.2, 5.0 Hz, 3H, Θ CH₃); ¹³C NMR (126 MHz, CDCl₃): δ 166.75, 150.34, 137.75, 135.05, 130.75, 124.33, 121.33, 112.68, 112.00, 60.73, 14.26; ESI-MS: m/z [M+H]⁺ 249.

Methyl(*E*)-3-(2-hydroxy-5-nitrophenyl)acrylate, 3s: Colourless solid. m.p. 131-133°C. ¹H NMR (500 MHz, CDCl₃): δ 8.41 (d, J = 2.7 Hz, 1H, aromatic), 8.15 (dd, J = 8.9, 2.7 Hz, 1H, aromatic), 7.95 (d, J =

16.1 Hz, 1H, CH), 6.94 (d, $J = 8.9$ Hz, 1H, aromatic), 6.70 (d, $J = 16.2$ Hz, 1H, CH), 3.85 (s, 3H, OCH₃); ESI-MS: m/z [M+H]⁺ 224.

Ethyl (E)-3-(2-hydroxy-5-nitrophenyl)acrylate, 3t: Colourless solid. m.p.140-143°C. ¹H NMR (400 MHz, CDCl₃): 8.93 (s, 1H, OH), 8.42 (d, $J = 2.7$ Hz, 1H, aromatic), 8.15 (dd, $J = 8.9, 2.7$ Hz, 1H, aromatic), 8.05 (d, $J = 16.3$ Hz, 1H, CH), 7.00 (d, $J = 9.0$ Hz, 1H, aromatic), 6.81 (d, $J = 16.3$ Hz, 1H, CH), 4.35 (q, $J = 7.1$ Hz, 2H, OCH₂), 1.40 (t, $J = 7.1$ Hz, 3H, CH₃); ESI-MS: m/z [M+H]⁺ 238.

Methyl (E)-3-(3-bromo-2-hydroxy-5-nitrophenyl)acrylate, 3u: Colourless solid. m.p.140-142°C. ¹H NMR (500 MHz, CDCl₃): δ 8.41 (d, $J = 23.6$ Hz, 1H, aromatic), 8.00 (dd, $J = 80.4, 69.0$ Hz, 1H, aromatic), 6.68 (d, $J = 16.0$ Hz, 1H, CH), 4.43 (d, $J = 16.0$ Hz, 1H, CH), 3.81 (s, 3H, OCH₃); ESI-MS: m/z [M+H]⁺ 302.

Ethyl (E)-3-(3-bromo-2-hydroxy-5-nitrophenyl)acrylate, 3v: Colourless solid. m.p.149-151°C. ¹H NMR (500 MHz, CDCl₃): δ 8.41 (d, $J = 18.6$ Hz, 2H, aromatic), 7.94 (d, $J = 16.1$ Hz, 1H, CH), 6.68 (d, $J = 16.1$ Hz, 1H, CH), 4.30 (dd, $J = 13.8, 6.8$ Hz, 2H, OCH₂), 1.36 (t, $J = 6.9$ Hz, 3H, CH₃); ESI-MS: m/z [M+H]⁺ 316.

Methyl (E)-3-(2-hydroxy-3,5-dinitrophenyl)acrylate, 3w: Colourless solid. m.p.148-150°C. ¹H NMR (400 MHz, CDCl₃): δ 11.72 (s, 1H, OH), 9.07 (d, $J = 2.7$ Hz, 1H, aromatic), 8.68 (d, $J = 2.7$ Hz, 1H, aromatic), 7.98 (d, $J = 16.2$ Hz, 1H, CH), 6.76 (d, $J = 16.2$ Hz, 1H, CH), 3.86 (s, 3H, OCH₃); ¹³C NMR (101 MHz, CDCl₃): δ 166.22, 157.29, 139.75, 135.03, 133.27, 129.51, 127.64, 124.01, 121.90, 52.20; ESI-MS: m/z [M+H]⁺ 269.

Ethyl (E)-3-(2-hydroxy-3,5-dinitrophenyl)acrylate, 3x: Colourless solid. m.p.143-147°C. ¹H NMR (400 MHz, CDCl₃): δ 11.72 (s, 1H, OH), 9.07 (d, $J = 2.7$ Hz, 1H, aromatic), 8.68 (d, $J = 2.7$ Hz, 1H, aromatic), 7.97 (d, $J = 16.2$ Hz, 1H, CH), 6.76 (d, $J = 16.2$ Hz, 1H, CH), 4.32 (q, $J = 7.1$ Hz, 2H, OCH₂), 1.37 (t, $J = 7.1$ Hz, 3H, CH₃); ¹³C NMR (101 MHz, CDCl₃): δ 165.76, 157.27, 139.74, 134.72, 133.25, 129.45, 127.74, 124.48, 121.82, 61.16, 14.22; ESI-MS: m/z [M+H]⁺ 283.

DPPH free radical scavenging assay

Assay for the scavenging of stable free radical 1,1-diphenyl-2-picrylhydrazyl (DPPH) was done. Briefly, in a 96-well micro plate, 25 μ L of test sample dissolved in DMSO (1 mg/mL), 100 μ L of 0.1 M tris-

HCl buffer (pH 7.4) and 125 μ L of 0.5 mM DPPH solution dissolved in absolute ethyl alcohol were added. The reaction mixture was shaken well and incubated in dark for 30 min and read at 517 nm spectrophotometrically (Spectra Max plus 384, Molecular Devices Corporation, Sunnyvale, CA, USA). Percentage of DPPH scavenging was calculated as $(1-B/A) \times 100$ where A represents absorbance of control without test samples and B represents absorbance in presence of test samples.

ABTS⁺ free radical scavenging assay

Scavenging of the ABTS⁺ [2,2'-azino-bis(3-ethylbenzothiazoline-6-sulphonic acid)] cation was performed as described by Walker and Everette⁹. Briefly, 100 mL stock solution of ABTS⁺ (0.5 mM) was prepared by addition of 1 mL potassium persulfate (6.89 mM PBS, pH 8.0). The mixture was stored in the dark for 16 h. Test compounds were dissolved in DMSO (5mg/mL). Primary screening was done by mixing 10 μ L of test compounds in 100 μ L of methanol followed by 190 μ L of ABTS⁺ in a 96-well microplate. Absorbance of decolorized ABTS⁺ was measured at 734 nm after 15 min incubation in the dark on a BioTeksynergy⁴ multi-mode microplate reader. For each test sample a separate blank sample (devoid of ABTS⁺) was used for background subtraction. The percentage of ABTS⁺ scavenging was calculated applying following formula; % ABTS⁺ scavenging = [(Absorbance control-Absorbance test)/Absorbance control \times 100]. Various serial dilutions of active compounds were prepared and tested for determination of SC₅₀ values. Suitable regression analysis was applied for calculation of SC₅₀.

α -Glucosidase inhibitory assay

α -Glucosidase inhibitory activity was determined as per our earlier reported method. Rat intestinal acetone powder in normal saline (100:1; w/v) was sonicated properly and the supernatant was used as a source of crude intestinal α -glucosidase after centrifugation. In brief, 10 μ L of test samples (5 mg/mL DMSO solution) were reconstituted in 100 μ L of 100 mM-phosphate buffer (pH 6.8) in 96-well microplate and incubated with 50 μ L of crude intestinal α -glucosidase for 5 min before 50 μ L substrate (5 mM, *p*-nitrophenyl- α -D-glucopyranoside prepared in same buffer) was added. Release of *p*-nitrophenol was measured after 15 min incubation at

405 nm spectrophotometrically (SpectraMaxplus384), Molecular Devices Corporation, Sunnyvale, CA, USA) 5 min after incubation with substrate. Individual blanks for test samples were prepared to correct background absorbance where substrate was replaced with 50 μ L of buffer. Control sample contained 10 μ L DMSO in place of test samples. Percentage of enzyme inhibition was calculated as $(1 - B/A) \times 100$ where [A] represents absorbance of control without test samples, and [B] represents absorbance in the presence of test samples.

Conclusions

In conclusion, α,β -unsaturated esters **3a-x** have been prepared by the reaction of salicylaldehydes **1a-e** with Wittig reagents **2a-b** in dry DCM at RT. The compounds were evaluated for their free-radical scavenging and α -glucosidase inhibitory activities. Compounds **3c** and **3d** identified as DPPH free radical scavengers. Twenty compounds **3a-r** and **3u-v** have shown promising ABTS⁺ free radical scavenging activity. Compound **3r** identified as potent α -glucosidase inhibitor and compounds **3g** and **3p** have shown moderate α -glucosidase inhibitory activity.

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